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Synthesis of Novel Nucleosides with a Fused Cyclopropane Ring Substituted by a Hydroxymethyl Group

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Abstract—The two diastereomers of a 3-oxabicyclo[3.1.0]hexan-2-ol **21a**–**b** with the cyclopropane ring substituted by a hydroxymethyl group were synthesized from both products **17a** and **18a** obtained from the bromohydroxylation of epoxide **16a** derived from 3-oxabicyclo[3.2.0]hept-6-en-2-one **15**. This preparation involved two stereospecific C_4-C_3 ring contractions leading to *cis,cis*-trisubstituted cyclopropane compounds **18a** and **12**. The hydroxylactols **21a**–**b** thus obtained were diacetylated and the single product **22** was subjected to substitution at the anomeric position by protected or free bases, providing, in each case, only one of the possible products. Deprotection with ammonia yielded new bicyclic nucleosides **11a**–**d**. © 2000 Elsevier Science Ltd. All rights reserved.

Introduction

In recent years, numerous nucleosides with modifications of the sugar unit have been described with the hope of obtaining new antiviral and anticancer agents.¹ In this area, several works were devoted to the preparation of bicyclic compounds,² including cyclopropano homologs³ **2**, **3**, **4** of 2',3'-didehydro-2',3'-dideoxythymidine (d4T) **1**, which is one of the nucleosides approved by the F.D.A. for treatment of HIV infection. In these molecules, the double bond was replaced by a cyclopropane ring via cyclopropanation,^{3a} homologous Ferrier reaction,^{3b,c} 1,3-dipolar cycloaddition^{3d}



Figure 1.

Keywords: ring contraction; stereoselection; substitution; bicyclic nucleosides.

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Scheme 1.

or Michael addition.^{3e} These analogs were expected to be more stable than 1 to glycosyl cleavage because the intermediate carbonium ion should be less stabilized.^{3b} As such a cleavage leads to inactivation, this modification should be interesting if these compounds keep the biological properties. Unfortunately, **3b** had only a weak anti-HIV activity^{3c,d} and **3a** was inactive against several viruses.^{3b} Other bicyclonucleosides disubstituted⁴ on the cyclopropane moiety **5–7**, and tri- or tetrasubstituted^{3e,5} **8–10** have also been prepared (Fig. 1) for potential applications in the field of therapeutics based on nucleosides or oligonucleotides.

In the course of our research program concerning the synthesis of new carbocyclic nucleosides, we obtained the lactone 13,⁶ in several steps, starting from cyclobutene anhydride 14.7 We intended to use it in the preparation of the corresponding non-halogenated diol by concomitant reduction of the lactone moiety and of the carbon-halide bond. However, reaction with LiAlH₄ proceeded without removal of the halide and a following reduction of this halogenated diol in more drastic conditions (LiEt₃BH) produced a certain amount of the unexpected ring contraction product 12.⁶ Nevertheless, we thought that this compound 12, with a cis, cis-relationship between substituents of the cyclopropane moiety, which would probably not be easily available by another way, might be interesting for synthetic applications. We thus envisaged improving the experimental conditions for obtaining 12 and then using it for the preparation of new fused cyclopropane-containing nucleosides 11 by substitution at the anomeric position (Scheme 1).

Results and Discussion

In previous works, anhydride 14^7 was reduced to the lactone 15^8 that was epoxidized leading to 16a, predominantly.^{8c} Treatment of epoxide 16a with aqueous hydrobromic acid afforded a mixture of bromohydrin 17a and compound 18a, the configuration of which was not assigned.^{8c} We have now improved both their yields and this aldehyde 18a has been identified as the *cis,cis*-isomer (see below). At this point, to gain insight into the mechanism of the ring contraction that occurred in acidic medium, some additional experiments were carried out. We thus subjected epoxide 16b to the same experimental conditions and also obtained a mixture of two compounds, bromohydrin 19a and aldehyde 18b (Scheme 2).

Identification of **19a** began by assignment of H-5 owing to its coupling together with H-1, H-4, H-6 and even with H-7 (0.8 Hz). Successive spin decoupling experiments starting from H-5 led to assignments of H-1, H-4, H-6 and H-7. Configurations were then deduced from NOE experiments (Fig. 2). The structures of aldehydes **18a** and **18b** were also based on NOE experiments and on the fact that the *cis* vicinal coupling constants are always larger than the *trans* ones for two stereomer cyclopropanes.⁹ Furthermore, the relative instability of **18a**, which provided **20** slowly on standing, also confirmed the *cis,cis*-stereochemistry of **18a** (Scheme 3).

Ring contractions from epoxides and bromohydrins have already been reported by numerous research groups and





Figure 2. NOE enhancements (%) and coupling constants.



Scheme 3.

have led to several synthetic applications.^{6,10} In the case of the formation of **18a** and **18b**, the interesting point is obtaining only one isomer in each reaction. When starting from **16a**, bromohydrin **17a** was formed and appeared relatively stable in the reaction mixture. As the other isomer **17b** was not observed, one hypothesis for the formation of **18a** would be via the ring contraction from this bromohydrin. However, it is more likely that **17b** is not obtained at all but that **18a** is derived directly from an intermediate carbonium ion stabilized by anchimeric assistance of the vicinal carbonyl group (Scheme 4). Breaking the C–C bond close to the hydroxyl group and attacking the corresponding electron doublet from the other side could thus provide the rearranged carbonium ion with the *cis,cis*-structure. In contrast, for the reaction from **16b**, protonation leads to an intermediate with the hydroxyl group on the same side as the carbonyl



Scheme 4.





Scheme 6. (a) 1: BSA, MeCN; 2: TMSOTf; 3: 22, 2 h; 4: NaHCO₃; (b) NH₃, MeOH, 2-7 days.

group and such an anchimeric stabilization should be more difficult. Therefore, in this case, the ring contraction probably works via bromohydrin **19b**, which may explain obtaining the *trans* product.^{10c} Finally, in both cases, bromohydrins **17a** or **19a**, with the hydroxyl group linked to C-6, could be isolated, whereas the 7-hydroxy intermediates yielded the cyclopropane compounds **18a** and **18b**.

Subsequently, we tried to optimize the synthesis of compound 12 with respect to our previous report.⁶ After protection of bromohydrin 17a with a silyl group to afford the lactone 13,⁶ we were pleased to observe that 12 could be directly prepared from 13 in one step instead of two in the early experiment. As a matter of fact, when 13 was reduced by 5 molar equivalents of LiEt₃BH in THF, an 85% isolated yield of 12 was obtained (Scheme 5). The subsequent desilylation afforded the lactol 21 in a good yield as a

mixture of both epimers in a 92:8 ratio. To avoid the protection-deprotection sequence, we also tried to subject **17a** to the same experimental conditions but we only obtained degradation products. Protection of the hydroxyl group then appeared as necessary. On the other hand, attempts of ring contraction of **17a** to **18a**, in basic medium and in several experimental conditions,¹⁰ led either to recovery of the starting material or to degradation products.

These improvements to prepare 12, 17a and 18a were important on the synthetic point of view, especially as we envisaged using not only 12, derived from 17a, but also 18a, to produce compound 21, the key intermediate of this work. Eventually reduction of 18a by Dibal-H¹¹ afforded the lactol 21 in a satisfying yield (71%). Then acetylation led to the crude diacetate 22 as a single diastereomer. Purification by flash chromatography provided 22 in an 80% isolated yield



Figure 3. NOE enhancements (%).

together with a certain amount of the hydrolysis product at the anomeric position (10%, see the Experimental section). Configuration of the anomeric carbon was deduced from the 4.3% NOE enhancement of H-2 upon saturation of H-7. Compound **22** was thus available in satisfying overall yield (43% from **16a** when using **17a** and **18a**). This good result encouraged us to carry on our project towards the synthesis of bicyclic nucleosides.

When diacetate **22** was subjected to the one-pot substitution by thymine in Vorbrüggen et al. conditions,¹² modified by Dudycz et al.¹³ (BSA, TMSOTf, MeCN), the expected product **23a** was obtained in excellent yield (Scheme 6). Condensation also worked with protected purine bases but in lower yield (63–68%). All attempts to increase the sugarbase coupling yield (amount of catalyst, of silylating agent, temperature, solvent composition...) have been unsuccessful



and predominantly led to byproducts. However, regioselectivity was excellent from benzoyladenine, which exclusively led to the N-9 isomer **23b**. On the other hand, reaction with acetyl guanine gave a 1.5:1 mixture of the N-9/N-7 products, **23c** and **23d**, respectively, which were fortunately separable by column chromatography. As observed by several groups,¹⁴ the use of disubstituted guanine **24** is necessary to overcome this problem and to get exclusively the N-9 derivative. In a last step, bicyclic nucleosides **11** were obtained after deprotection with a solution of ammonia in methanol.¹⁵

Stereochemical assignments were deduced from NOE experiments for **11a**, **11b**, **11c** and **23d** (Fig. 3). In each case, significant enhancement of signal for the anomeric proton H-2' upon saturation of H-7' (5.9-8.4%) showed that the nucleobases were fixed at the opposite side from the acetoxymethyl or hydroxymethyl group. Obtaining only one anomer may be due to the steric hindrance of the acetoxymethyl group.

Regiochemistries were elucidated by HMBC experiments (Fig. 4). In the case of 23b, one quaternary carbon was correlated with H-2 and H-8, another one with H-8 and the third one with H-2. As C-5 is always at a higher field than C-4 and C-6 in adenine derivatives, H-2 and H-8 were thus distinguished. A ${}^{1}J^{13}C/{}^{1}H$ correlation then led to assignments of C-8 and C-2. Finally, the long range correlations between H-2' and C-8 and C-4 proved the N-9 regiochemistry. In the case of 11c and 11d, H-8 was correlated only with C-4 and C-5 and as C-5 is at a higher field than C-4, they were both assigned. The long range correlations between H-2' and C-8 and C-4 proved the N-9 regiochemistry for 11c. For the nucleoside 11d, correlation between H-2' and C-5 could not been observed, but as 11c had been undoubtedly identified, the H-2'/C-8 correlation proved the N-7 regiochemistry. Moreover, obtaining 11c from 23e was in full agreement with the previous results.

In conclusion, we pointed out that diacetate 22 was efficiently synthesized from the easily available epoxide 16a, and we also proposed interpretations of our results of ring contractions. New bicyclic nucleosides 11a-d were afterwards obtained from 22 by substitution at the anomeric position followed by deprotection. Biological tests showed that these compounds did not have antitumor properties. Their evaluations as anti-HIV agents are in progress. One of the interests of this strategy is the possibility of obtaining bicyclic compounds with a fused functionalized cyclopropane. However, coupling of bases exclusively led to products with an α -type configuration at the anomeric position. The use of this new route to prepare compounds with hydroxymethyl substituents linked to the five membered ring, which should have a better chance of having biological properties, is in due course.

Experimental

General

All the moisture-sensitive reactions were carried out in

oven-dried glassware (110°C) and under nitrogen atmosphere. Commercially available reagents and solvents were purified and dried, when necessary, by standard methods just prior to use. IR spectra were scanned on a FT infrared spectrophotometer. All melting points are uncorrected. ¹H and ¹³C NMR spectra were recorded at 400 and 100.6 MHz, respectively. Chemical shifts are reported in ppm downfield from TMS which was used as an internal reference. Multiplicities in the ¹³C spectra were determined by DEPT experiments, and numerous assignments were obtained by ¹³C/¹H HETCOR and HMBC experiments. Ratios in mixtures of epimers were calculated from ¹H NMR. Elemental analyses were obtained from the Service de Microanalyse, CNRS ICSN, Gif-sur-Yvette. High-resolution mass measurements were performed at the CRMPO, Rennes.

 $(1R^*, 5S^*, 6S^*, 7S^*)$ -7-Bromo-6-hydroxy-3-oxabicyclo[3.2.0]heptan-2-one (17a) and $(1S^*, 5S^*, 6R^*)$ -6-carbaldehyde-3oxabicyclo[3.1.0]hexan-2-one (18a) and compound 20. To an ice-cold solution of epoxide $16a^{8c}$ (2.07 g, 16.41 mmol) in acetone (130 mL) was added dropwise HBr (48% aqueous solution, 5.60 mL, 49.50 mmol). The reaction mixture was stirred for 7 h at room temperature and then was neutralized with an aqueous saturated solution of NaHCO₃ (40 mL). Acetone was removed in vacuo and the aqueous layer was extracted with EtOAc (6×40 mL). The combined organic layers were dried over MgSO4 and concentrated under reduced pressure. Purification of the crude product by column chromatography (silica gel, CH₂Cl₂/Et₂O: $1/0 \rightarrow 7/3$) afforded compound **18a** (0.65 g, 5.15 mmol, 31%) as colorless crystals and then compound 17a (1.60 g, 7.73 mmol, 47%) as white crystals.

Compound 17a. see Ref. 8c.

Compound **18a**. mp 55–56°C; IR (KBr, cm⁻¹) 1762, 1698, 1691, 1168; ¹H NMR (CDCl₃) δ 9.47 (d, 1H, CHO, J=5.2 Hz), 4.61 (dd, 1H, H-4, J=10.5, 5.0 Hz), 4.51 (ddd, 1H, H-4', J=10.5, 0.8, 0.8 Hz), 2.84 (m, 1H, H-5), 2.70 (ddd, 1H, H-1, J=8.5, 6.0, 0.8 Hz), 2.37 (ddd, 1H, H-6, J=8.5, 7.9, 5.2 Hz); ¹³C NMR (CDCl₃) δ 196.4 (CHO), 172.2 (C=O), 66.7 (CH₂), 30.6 (CH), 27.3 (CH), 25.6 (CH); Anal. Calcd for C₆H₆O₃ (MW: 126.11): C, 57.14; H, 4.79. Found: C, 57.04; H, 4.81.

Compound **20**. **18a** gave polycyclic compound **20** as a colorless oil, when it was left at room temperature for several days: IR (neat, cm⁻¹) 1764, 1191, 929; ¹H NMR (CDCl₃) δ 6.06 (d, 1H, *J*=3.3 Hz), 4.39 (dd, 1H, *J*=10.0, 5.2 Hz), 4.07 (dd, 1H, *J*=10.0, 1.4 Hz), 3.46 (ddd, 1H, *J*=7.3, 5.6, 3.3 Hz), 2.50 (m, 1H), 2.35 (dd, 1H, *J*=7.9, 5.6 Hz); ¹³C NMR (CDCl₃) δ 173.6 (*C*=O), 105.4 (*C*H), 68.7 (*C*H₂), 36.2 (*C*H), 30.4 (*C*H), 27.8 (*C*H); HRMS Calcd for C₆H₆O₃: (M+H)⁺ 127.0395. Found: 127.0394.

(1*S**,5*S**,6*S**)-6-Carbaldehyde-3-oxabicyclo[3.1.0]hexan-2-one (18b) and (1*R**,5*S**,6*R**,7*R**)-7-bromo-6-hydroxy-3-oxabicyclo[3.2.0]heptan-2-one (19a). Following the same procedure as for the preparation of 17a and 18a, epoxide 16b (0.40 g, 3.17 mmol) gave, after chromatography (silica gel, CH₂Cl₂/Et₂O: $1/0 \rightarrow 4/1$), compound 18b (50 mg, 0.40 mmol, 13%) as a colorless oil and compound 19a (268 mg, 1.29 mmol, 41%) as white crystals.

Compound **18b.** IR (neat, cm⁻¹) 1768, 1712, 1172; ¹H NMR (CDCl₃) δ 9.62 (d, 1H, CHO, J=2.7 Hz), 4.48 (dd, 1H, H-4, J=10.1, 4.5 Hz), 4.36 (dd, 1H, H-4', J=10.1, 0.6 Hz), 2.78 (ddd, 1H, H-5, J=6.3, 4.5, 2.7 Hz), 2.70 (ddd, 1H, H-1, J=6.3, 2.7, 0.6 Hz), 2.33 (ddd, 1H, H-6, J=2.7, 2.7, 2.7 Hz); ¹³C NMR (CDCl₃) δ 195.4 (CHO), 173.0 (C=O), 68.9 (CH₂), 32.4 (CH), 26.3 (CH), 25.9 (CH); HRMS Calcd for C₆H₆O₃: 126.0317. Found: 126.0314.

Compound **19***a*. mp 94–95°C (Et₂O); IR (KBr, cm⁻¹) 3400, 1743, 1192; ¹H NMR (acetone- d_6) δ 5.52 (br s, 1H, OH), 4.74 (dd, 1H, H-4, J=9.8, 4.6 Hz), 4.63 (ddd, 1H, H-6, J=8.0, 5.8, 0.7 Hz), 4.41 (dd, 1H, H-4', J=9.8, 9.8 Hz), 4.36 (ddd, 1H, H-7, J=5.8, 5.8, 0.8 Hz), 3.63 (m, 1H, H-5), 3.04 (ddd, 1H, H-1, J=8.4, 5.8, 0.7 Hz); ¹³C NMR (acetone- d_6) δ 176.1 (*C*=O), 74.1 (*C*H), 66.2 (*C*H₂), 49.7 (*C*H), 43.0 (*C*H), 39.8 (*C*H); Anal. Calcd for C₆H₇O₃Br (MW: 207.02): C, 34.81; H, 3.41; Br, 38.60. Found: C, 34.74; H, 3.41; Br, 38.52.

(1S^{*},2R^{*},5R^{*},6S^{*})-6-Hydroxymethyl-2-[(*tert*-butyldimethylsilyl)oxy]-3-oxabicyclo[3.1.0]hexane (12). A solution of 13 (2.73 g, 8.50 mmol) in anhydrous THF (84 mL) was added dropwise to a solution of LiEt₃BH (42.50 mL of a 1 M solution in THF, 42.50 mmol). The resulting mixture was stirred for 5 days at reflux. After cooling to 0°C, KOH 2 M (4.25 mL, 8.50 mmol) then H₂O (4 mL) were added and the solvent was evaporated. The residue was dissolved in CH₂Cl₂ (500 mL), washed (brine, 140 mL), dried over MgSO₄ and concentrated in vacuo. Purification by column chromatography (silica gel, $CH_2Cl_2/EtOAc: 1/0 \rightarrow 4/1$) afforded 12 (1.76 g, 7.22 mmol, 85%) as a colorless oil: IR (neat, cm⁻¹) 3350, 2929, 2856, 1251, 1083, 1039, 836; ¹H NMR (CDCl₃) δ 5.32 (s, 1H, H-2), 4.14 (dd, 1H, H-4, J=8.7, 3.7 Hz), 3.82 (d, 1H, H-4', J=8.7 Hz), 3.74 (d, 2H, CH_2OH , J=7.5 Hz), 1.86 (m, 1H, H-5), 1.80 (dd, 1H, H-1), J=8.1, 7.0 Hz), 1.34 (br s, 1H, OH), 1.22 (m, 1H, H-6), 0.90 (s, 9H, (CH₃)₃C), 0.12 (s, 3H, CH₃Si), 0.11 (s, 3H, CH₃Si); ¹³C NMR (CDCl₃) δ 97.8 (C2), 66.4 (C4), 57.4 (CH₂OH), 28.5 (C1), 25.7 ((CH₃)₃C), 21.2 (C6), 19.9 (C5), 17.9 ((CH₃)₃C), -4.3 (CH₃Si), -5.0 (CH₃Si); Anal. Calcd for C₁₂H₂₄O₃Si (MW: 244.41): C, 58.97; H, 9.90; Si, 11.49. Found: C, 58.62; H, 9.91; Si, 11.74; HRMS Calcd for $C_{12}H_{24}O_3Si: (M-H)^+$ 243.1416. Found: 243.1422.

(1*S*^{*},2*S*^{*},5*R*^{*},6*S*^{*})-6-Hydroxymethyl-3-oxabicyclo[3.1.0]hexan-2-ol (21a) and (1*S*^{*},2*R*^{*},5*R*^{*},6*S*^{*})-6-hydroxymethyl-3-oxabicyclo[3.1.0]hexan-2-ol (21b). Procedure A: A 1 M solution of *n*-Bu₄NF in THF (16.00 mL, 16.00 mmol) was added to a solution of compound 12 (1.37 g, 5.60 mmol) in anhydrous THF (24 mL). The reaction mixture was stirred at room temperature for 5 h. After removal of the solvent under reduced pressure, the residue was purified by column chromatography (silica gel, EtOAc/ MeOH: 1/0→98/2) to afford a mixture of both epimers 21a and 21b in a 92:8 ratio (641 mg, 4.93 mmol, 88%) as a colorless oil: IR (neat, cm⁻¹) 3382, 2886, 1022; ¹H NMR major isomer 21a (acetone-*d*₆) δ 5.22 (d, 1H, H-2, *J*=5.7 Hz), 5.07 (d, 1H, OH, *J*=5.7 Hz), 4.04 (dd, 1H, H-4, J=8.4, 3.8 Hz), 3.73 (d, 1H, H-4', J=8.4 Hz), 3.59 (m, 2H, CH₂OH), 3.41 (t, 1H, CH₂OH, J=5.5 Hz), 1.77 (m, 1H, H-5), 1.69 (dd, 1H, H-1, J=8.3, 6.8 Hz), 1.17 (m, 1H, H-6); ¹H NMR minor isomer **21b** (acetone- d_6) 5.80 (d, 1H, OH, J=9.8 Hz), 5.49 (dd, 1H, H-2, J=9.8, 2.8 Hz), 4.37 (dd, 1H, CH₂OH, J=5.3, 4.8 Hz), 4.00 (ddd, 1H, CH₂OH, J=11.3, 7.4, 4.8 Hz), 3.89 (dd, 1H, H-4, J=9.1, 3.3 Hz), 3.86 (d, 1H, H-4', J=9.1 Hz), 3.68 (ddd, 1H, CH₂OH, J=11.3, 10.1, 5.3 Hz), 1.80 (ddd, 1H, H-1, J=8.1, 7.2, 2.8 Hz), 1.26 (m, 1H, H-6) (H-5 hidden by a signal of the major isomer); ¹³C NMR major isomer **21a** (acetone- d_6) δ 97.9 (CH), 66.4 (CH₂), 56.9 (CH₂), 28.1 (CH), 22.4 (CH), 20.5 (CH); ¹³C NMR minor isomer **21b** (acetone- d_6) δ 100.6 (CH), 66.0 (CH₂), 57.1 (CH₂), 26.2 (CH), 22.3 (CH), 21.4 (CH); Anal. Calcd for C₆H₁₀O₃·0.2H₂O (MW: 133.75): C, 53.88; H, 7.84. Found: C, 53.92; H, 7.84.

Procedure B: To a cold solution (-78°C) of **18a** (314 mg, 2.49 mmol) in anhydrous THF (5.4 mL) was added a 1 M solution of Dibal-H in THF (7.60 mL, 7.60 mmol) over a period of 1 h and in a temperature below -68°C . After 2 h stirring at -78°C , the reaction mixture was quenched with MeOH (1.4 mL) and allowed to warm to room temperature. EtOAc (14 mL) and saturated NaHCO₃ solution (1.4 mL) were added and the mixture was stirred for 2 h. Powdered Na₂SO₄ (3.2 g) was then added and stirring was continued overnight. The precipitate was removed by filtration and washed with EtOAc. The solvent was evaporated in vacuo and the residue thus obtained was purified by column chromatography (silica gel, EtOAc/CH₂Cl₂: $1/1 \rightarrow 2/1 + 5\%$ MeOH) to yield a mixture of both epimers **21a** and **21b** in a 92:8 ratio (232 mg, 1.78 mmol, 71%) as a colorless oil.

(1*R**,2*R**,5*R**,6*S**)-2-Acetyloxy-6-acetyloxymethyl-3-oxabicyclo[3.1.0]hexane (22). Acetic anhydride (1.70 mL, 18.10 mmol) was added dropwise to an ice-cold stirred solution of 21a+21b (460 mg, 3.53 mmol) in pyridine (5.0 mL). The reaction mixture was stirred for 30 min at 0°C and then at room temperature overnight. After cooling to 0°C, MeOH (2.0 mL) was added. The mixture was then allowed to warm to room temperature and concentrated to dryness. The crude product was purified by flash chromatography (silica gel, cyclohexane/EtOAc: 9/1 \rightarrow 2/1) to yield diacetate 22 (604 mg, 2.82 mmol, 80%) as a colorless oil and lactol obtained from partial hydrolysis of 22 on silica gel (61 mg, 0.35 mmol, 10%, 97:3 ratio of epimers) as a colorless oil.

Compound 22. IR (neat, cm⁻¹) 1741, 1236; ¹H NMR (CDCl₃) δ 6.18 (s, 1H, H-2), 4.23 (dd, 1H, CH₂OAc, J=11.7, 7.7 Hz), 4.20 (dd, 1H, H-4, J=9.0, 3.4 Hz), 4.13 (dd, 1H, CH₂OAc, J=11.7, 7.9 Hz), 3.97 (d, 1H, H-4', J=9.0 Hz), 2.08 (s, 3H, CH₃), 2.07 (s, 3H, CH₃), 1.99–1.90 (m, 2H, H-1 and H-5), 1.41 (m, 1H, H-6); ¹³C NMR (CDCl₃) δ 170.9 (C=O), 170.1 (C=O), 97.4 (CH), 67.9 (CH₂), 59.3 (CH₂), 25.8 (CH), 21.2 (CH₃), 20.9 (CH₃), 19.6 (CH), 17.7 (CH); Anal. Calcd for C₁₀H₁₄O₅ (MW: 214.22): C, 56.07; H, 6.59. Found: C, 55.99; H, 6.69.

Hydrolysis product. IR (neat, cm⁻¹) 3425, 1732, 1240; ¹H NMR major isomer (CDCl₃) δ 5.35 (d, 1H, H-2, *J*=4.3 Hz), 4.25–4.12 (m, 3H, CH₂OAc and H-4), 3.89 (d, 1H, H-4', *J*=8.9 Hz), 2.88 (d, 1H, OH, *J*=4.3 Hz), 2.07 (s, 3H, CH₃),

1.95–1.86 (m, 2H, H-1 and H-5), 1.34 (m, 1H, H-6); ¹H NMR minor isomer (CDCl₃) δ 5.60 (dd, 1H, H-2, J=10.5, 2.6 Hz), 4.60 (dd, 1H, CH₂OAc, J=12.0, 6.0 Hz), 4.45 (dd, 1H, CH₂OAc, J=12.0, 9.2 Hz), 4.36 (d, 1H, OH, J=10.5 Hz), 3.99 (dd, 1H, H-4, J=9.5, 3.2 Hz), 3.96 (d, 1H, H-4', J=9.5 Hz), 2.10 (s, 3H, CH₃) 1.86–1.79 (m, 2H, H-1 and H-5) (H-6 hidden by a signal of the major isomer); ¹³C NMR major isomer (CDCl₃) δ 171.1 (*C*=O), 97.4 (CH), 66.4 (CH₂), 59.8 (CH₂), 27.2 (CH), 21.0 (CH₃), 19.8 (CH), 17.8 (CH); HRMS Calcd for C₈H₁₂O₄: 172.0735. Found: 172.0737.

 $(1'R^*, 2'S^*, 5'R^*, 6'S^*)$ -1-(6'-Acetyloxymethyl-3'-oxabicyclo-[3.1.0]hex-2'-yl)thymine (23a). BSA (306 µL, 1.20 mmol) was added to a suspension of thymine (53 mg, 0.42 mmol) in anhydrous MeCN (3 mL), and stirring was continued for 15 min at room temperature until a clear solution was obtained. Diacetate 22 (86 mg, 0.40 mmol) in anhydrous MeCN (2 mL) and TMSOTf (80 µL, 0.41 mmol) were successively added at 0°C. The reaction mixture was stirred at room temperature for 2 h. CH₂Cl₂ (2 mL) was added and the reaction mixture was quenched with aqueous saturated $NaHCO_3$ (4.0 mL). Solvents were removed in vacuo, the residue was dissolved in EtOAc (25 mL) and the solution was washed with water (20 mL), dried over MgSO₄ and evaporated to dryness. The crude product was purified by column chromatography (silica gel, CH₂Cl₂/EtOAc: $1/0 \rightarrow 2/1$) to give 23a (101 mg, 0.36 mmol, 90%) as a white solid: mp 211–212°C; IR (KBr, cm⁻¹) 3035, 1724, 1704, 1662, 1249; ¹H NMR (CDCl₃) δ 8.95 (br s, 1H, NH), 7.11 (d, 1H, H6, J=0.9 Hz), 6.08 (s, 1H, H-2'), 4.28 (dd, 1H, H-4', J=9.3, 4.0 Hz), 4.26 (dd, 1H, CH₂OAc, J=11.9, 7.7 Hz), 4.20 (dd, 1H, CH_2OAc , J=11.9, 7.7 Hz), 4.05 (d, 1H, H-4", J=9.3 Hz), 2.20 (m, 1H, H-5'), 2.10 (s, 3H, CH₃CO₂), 2.03 (dd, 1H, H-1', J=8.4, 6.6 Hz), 1.95 (d, 3H, CH₃ base, J=0.9 Hz), 1.57 (m, 1H, H-6'); ¹³C NMR (CDCl₃) δ 171.0 (CH₃CO₂), 163.8 (C-4), 150.5 (C-2), 135.4 (C-6), 111.1 (C-5), 85.3 (C-2'), 68.9 (C-4'), 59.0 (CH₂OAc), 26.4 (C-1'), 21.8 (C-5'), 20.9 (CH₃CO₂), 18.4 (C-6'), 12.7 (CH₃) base); Anal. Calcd for C₁₃H₁₆N₂O₅ (MW: 280.28): C, 55.71; H, 5.75; N, 9.99. Found: C, 55.89; H, 5.76; N, 9.97; HRMS Calcd for C₁₃H₁₆N₂O₅: 280.1059. Found: 280.1057.

 $(1'R^*, 2'S^*, 5'R^*, 6'S^*)$ -9-(6'-Acetyloxymethyl-3'-oxabicyclo-[3.1.0]hex-2'-yl)-6-N-benzoyladenine (23b). Reaction from BSA (450 µL, 1.76 mmol), 6-N-benzoyladenine (200 mg, 0.84 mmol), diacetate 22 (123 mg, 0.57 mmol) and TMSOTf (145 µL, 0.75 mmol), in the same experimental conditions as for 23a, led to 23b as a white solid (142 mg, 0.36 mmol, 63%) after column chromatography (silica gel, CH₂Cl₂/MeOH: 98/2): mp 173-174°C; IR (KBr, cm⁻ 1726, 1705, 1514, 1248; ¹H NMR (CDCl₃) δ 9.06 (br s, 1H, NH), 8.81 (s, 1H, H-2), 8.14 (s, 1H, H-8), 8.03 (d, 2H, arom, J=7.3 Hz), 7.61 (m, 1H, arom), 7.53 (m, 2H, arom), 6.37 (s, 1H, H-2'), 4.39 (dd, 1H, CH₂OAc, J=11.8, 7.3 Hz), 4.33 (dd, 1H, H-4', J=9.3, 3.6 Hz), 4.31 (dd, 1H, CH₂OAc, J=11.8, 8.1 Hz), 4.13 (d, 1H, H-4", J=9.3 Hz), 2.37 (dd, 1H, H-1', J=8.1, 6.7 Hz), 2.31 (m, 1H, H-5'), 2.13 (s, 3H, CH₃), 1.65 (m, 1H, H-6'); ¹³C NMR (CDCl₃) δ 171.0 (CH₃CO), 164.7 (C₆H₅CO), 152.7 (C-2), 151.6 (C-4), 149.5 (C-6), 140.8 (C-8), 133.5 (arom C), 132.7 (arom C), 128.7 (arom C), 127.8 (arom C), 123.4 (C-5), 83.9 (C-2'), 68.4 (C-4'), 59.0 (CH₂OAc), 25.8 (C-1'), 21.8 (C-5'), 21.0 (CH₃),

18.7 (C-6'); Anal. Calcd for C₂₀H₁₉N₅O₄ (MW: 393.40): C, 61.06; H, 4.87; N, 17.80. Found: C, 61.14; H, 4.94; N, 17.63.

 $(1'R^*, 2'S^*, 5'R^*, 6'S^*)$ -9-(6'-Acetyloxymethyl-3'-oxabicyclo-[3.1.0]hex-2'-yl)-2-*N*-acetylguanine (23c) and $(1'R^*, 2'S^*, 5'R^*, 6'S^*)$ -7-(6'-acetyloxymethyl-3'-oxabicyclo[3.1.0]hex-2'-yl)-2-*N*-acetylguanine (23d). Compounds 23c and 23d were prepared as described for 23a using BSA (1.30 mL, 5.10 mmol), 2-*N*-acetylguanine (325 mg, 1.68 mmol), diacetate 22 (236 mg, 1.10 mmol) and TMSOTf (238 μ L, 1.23 mmol). Both compounds were separated by column chromatography (silica gel, CH₂Cl₂/MeOH: 99/1 \rightarrow 96/4) to give isomer 23d as a white solid (102 mg, 0.29 mmol, 27%) and then isomer 23c as a white powder (157 mg, 0.45 mmol, 41%).

Compound **23c**. mp 216–217°C (MeOH); IR (KBr, cm⁻¹) 3126, 1736, 1680, 1614, 1556, 1253; ¹H NMR (DMSOd₆) δ 12.04 (br s, 1H, H1), 11.71 (br s, 1H, NHAc), 8.16 (s, 1H, H-8), 6.06 (s, 1H, H-2'), 4.21 (dd, 1H, CH₂OAc, *J*=11.8, 7.2 Hz), 4.18–4.11 (m, 2H, CH₂OAc and H-4'), 3.94 (d, 1H, H-4", *J*=9.1 Hz), 2.34–2.25 (m, 2H, H-5' and H-1'), 2.18 (s, 3H, CH₃CON), 2.06 (s, 3H, CH₃CO₂), 1.54 (m, 1H, H-6'); ¹³C NMR (DMSOd₆) δ 173.5 (CH₃CON), 170.5 (CH₃CO₂), 154.8 (C-6 or C-2), 148.0 (C-4), 147.9 (C-2 or C-6), 137.4 (C-8), 120.2 (C-5), 82.3 (C-2'), 67.2 (C-4'), 59.0 (CH₂OAc), 25.0 (C-1'), 23.8 (CH₃CON), 21.2 (C-5'), 20.8 (CH₃CO₂), 17.8 (C-6'); Anal. Calcd for C₁₅H₁₇N₅O₅·0.3H₂O (MW: 352.73): C, 51.08; H, 5.03; N, 19.85. Found: C, 50.88; H, 4.91; N, 19.91.

Compound **23***d*. mp 119–120°C; IR (KBr, cm⁻¹) 3153, 1734, 1685, 1614, 1367, 1246; ¹H NMR (CDCl₃) δ 12.40 (br s, 1H, NH), 11.16 (br s, 1H, NH), 8.01 (s, 1H, H-8), 6.59 (s, 1H, H-2'), 4.35 (dd, 1H, CH₂OAc, *J*=11.9, 7.8 Hz), 4.30 (dd, 1H, CH₂OAc, *J*=11.9, 7.6 Hz), 4.26 (dd, 1H, H-4', *J*=9.4, 3.6 Hz), 4.15 (d, 1H, H-4'', *J*=9.4 Hz), 2.43 (s, 3H, CH₃CON), 2.32 (dd, 1H, H-1', *J*=8.3, 6.6 Hz), 2.21 (m, 1H, H-5'), 2.12 (s, 3H, CH₃CO2), 1.63 (m, 1H, H-6'); ¹³C NMR (CDCl₃) δ 173.3 (CH₃CON), 171.1 (CH₃CO₂), 156.9 (C-4), 153.2 (C-6 or C-2), 148.1 (C-2 or C-6), 140.3 (C-8), 111.3 (C-5), 85.9 (C-2'), 68.1 (C-4'), 59.0 (CH₂OAc), 27.0 (C-1'), 24.6 (CH₃CON), 21.2 (C-5'), 21.0 (CH₃CO₂), 18.6 (C-6'); Anal. Calcd for C₁₅H₁₇N₅O₅-0.3H₂O (MW: 352.73): C, 51.08; H, 5.03; N, 19.85. Found: C, 51.09; H, 4.96; N, 19.95.

 $(1^{\prime}R^{*}, 2^{\prime}S^{*}, 5^{\prime}R^{*}, 6^{\prime}S^{*})$ -9- $(6^{\prime}$ -Acetyloxymethyl-3^{\prime}-oxabicyclo-[3.1.0]hex-2'-yl)-2-N-acetyl-6-O-(diphenylcarbamoyl)guanine (23e). Reaction from BSA (356 µL, 1.40 mmol), 2-Nacetyl-6-O-(diphenylcarbamoyl)guanine (217 mg, 0.56 mmol), diacetate 22 (77 mg, 0.36 mmol) and TMSOTf $(80 \ \mu L, 0.41 \ mmol)$, in the same experimental conditions as for 23a, led to 23e as a white solid (118 mg, 0.22 mmol, 61%) after column chromatography (silica gel, CH₂Cl₂/MeOH: 99/1→96/4): mp 170–172°C; IR (KBr, cm⁻¹) 3276, 1736, 1284, 1188; ¹H NMR (CDCl₃) δ 8.10 (br s, 1H, NH), 8.04 (s, 1H, H8), 7.50-7.30 (m, 8H, arom), 7.26 (m, 2H, arom), 6.21 (s, 1H, H-2'), 4.37-4.31 (m, 2H, H-4' and CH₂OAc), 4.29 (dd, 1H, CH₂OAc, J=11.8, 8.2 Hz), 4.08 (d, 1H, H-4", J=9.2 Hz), 2.50 (s, 3H, CH₃), 2.33 (m, 1H, H-5'), 2.26 (dd, 1H, H-1', J=8.2, 6.7 Hz), 2.12 (s, 3H, CH₃), 1.61 (m, 1H, H-6⁷); ¹³C NMR (CDCl₃) δ 171.0 (C=O), 170.3 (C=O), 156.2 (quat C), 154.5 (quat C),

152.1 (quat C), 150.3 (quat C), 141.9 (*C*H), 141.7 (quat C), 129.2 (*C*H), 127.0 (quat C), 121.2 (quat C), 84.2 (*C*H), 68.7 (*C*H₂), 59.1 (*C*H₂), 25.9 (*C*H), 25.1 (*C*H₃), 22.0 (*C*H), 21.0 (*C*H₃), 18.7 (*C*H); HRMS Calcd for $C_{28}H_{26}N_6O_6$: (M+H)⁺ 543.1992. Found: 543.1988.

 $(1'S^*, 2'S^*, 5'R^*, 6'S^*)$ -1-(6'-Hydroxymethyl-3'-oxabicyclo-[3.1.0]hex-2'-yl)thymine (11a). Saturated NH₃ in MeOH (4 mL) was added with stirring to a solution of 23a (120 mg, 0.43 mmol) in MeOH (4 mL) at 0°C. The mixture was stirred at room temperature for 2 days and the solvent was evaporated in vacuo. Purification of the residue by column chromatography (silica gel, CH₂Cl₂/MeOH: 97/ $3\rightarrow 96/4$) gave the nucleoside 11a as a white powder (92 mg, 0.39 mmol, 90%): mp 202-204°C (MeOH), IR (KBr, cm⁻¹) 3469, 1691; ¹H NMR (DMSOd₆) δ 11.30 (br s, 1H, NH), 7.41 (d, 1H, H-6, J=1.0 Hz), 5.98 (s, 1H, H-2'), 4.60 (dd, 1H, OH, J=5.4, 5.4 Hz), 4.22 (dd, 1H, H-4', J=8.8, 4.1 Hz), 3.80 (d, 1H, H-4", J=8.8 Hz), 3.52–3.40 (m, 2H, CH₂OH), 2.17 (m, 1H, H-5'), 1.89 (dd, 1H, H-1', J=8.3, 6.9 Hz), 1.81 (d, 3H, CH₃, J=1.0 Hz), 1.30 (m, 1H, H-6'); 13 C NMR (DMSOd₆) δ 163.7 (C=O), 150.6 (C=O), 136.3 (C-6), 109.8 (quat C), 83.7 (C-2'), 68.1 (C-4'), 54.8 (*C*H₂OH), 25.5 (C-1[']), 21.6 (C-5[']), 21.3 (C-6[']), 12.2 (*C*H₃); Anal. Calcd for C₁₁H₁₄N₂O₄ (MW: 238.24): C, 55.46; H, 5.92; N, 11.76. Found: C, 55.48; H, 5.97; N, 11.74.

 $(1'S^*, 2'S^*, 5'R^*, 6'S^*)$ -9-(6'-Hydroxymethyl-3'-oxabicyclo-[3.1.0]hex-2'-yl)adenine (11b). Following the same procedure as for the preparation of 11a, compound 23b (117 mg, 0.30 mmol) gave, after column chromatography (silica gel, CH₂Cl₂/MeOH: $95/5 \rightarrow 90/10$), nucleoside **11b** as a white powder (69 mg, 0.28 mmol, 94%): mp 203°C dec. (MeOH); IR (KBr, cm⁻¹) 3357, 1662, 1597; ¹H NMR (DMSOd₆) δ 8.27 (s, 1H, H8), 8.15 (s, 1H, H-2), 7.29 (br s, 2H, NH₂), 6.20 (s, 1H, H-2'), 4.63 (dd, 1H, OH, J=5.5, 5.5 Hz), 4.20 (dd, 1H, H-4', J=8.7, 3.8 Hz), 3.90 (d, 1H, H-4", J=8.7 Hz), 3.63-3.52 (m, 2H, CH₂OH), 2.21 (m, 1H, H-5'), 2.14 (dd, 1H, H-1', J=8.1, 6.9 Hz), 1.37 (m, 1H, H-6'); ¹³C NMR (DMSOd₆) δ 156.0 (C-6), 152.7 (C-2), 148.9 (C-4), 138.7 (C-8), 118.8 (C-5), 82.3 (C-2'), 67.5 (C-4'), 54.8 (CH₂OH), 25.5 (C-1'), 21.8 (C-6'), 21.4 (C-5'); Anal. Calcd for C₁₁H₁₃N₅O₂ (MW: 247.26): C, 53.43; H, 5.30; N, 28.32. Found: C, 53.26; H, 5.23; N, 28.15.

 $(1'S^*, 2'S^*, 5'R^*, 6'S^*)$ -9-(6'-Hydroxymethyl-3'-oxabicyclo-[3.1.0]hex-2'-yl)guanine (11c). Saturated NH₃ in MeOH (4 mL) was added with stirring to a solution of 23c (103 mg, 0.30 mmol) in MeOH (4 mL) at 0°C. The mixture was stirred at room temperature for 2 days and the solvent was evaporated in vacuo. The residue was crystallized in MeOH/H₂O (5/1, 5 mL) to yield nucleoside 11c as a white powder (57 mg, 0.22 mmol, 73%): mp>315°C (MeOH/ H_2O ; IR (KBr, cm⁻¹) 3332, 1685, 1655; ¹H NMR (DMSOd₆) δ 10.60 (br s, 1H, H-1), 7.83 (s, 1H, H-8), 6.51 (br s, 2H, NH₂), 5.95 (s, 1H, H-2'), 4.59 (dd, 1H, OH, J=5.3, 5.3 Hz), 4.12 (dd, 1H, H-4', J=8.8, 3.8 Hz), 3.86 (d, 1H, H-4", J=8.8 Hz), 3.57 (ddd, 1H, CH₂OH, J=11.4, 7.1, 5.3 Hz), 3.49 (ddd, 1H, CH₂OH, J=11.4, 7.9, 5.3 Hz), 2.16 (m, 1H, H-5'), 2.05 (dd, 1H, H-1', J=8.2, 6.9 Hz), 1.33 (m, 1H, H-6'); ¹³C NMR (DMSOd₆) δ 156.8 (C-6 or C-2), 153.8 (C-2 or C-6), 150.6 (C-4), 134.9 (C-8), 116.5 (C-5), 81.7 (C-2'), 67.2 (C-4'), 54.8 (CH₂OH), 25.3

(C-1'), 21.6 (C-6'), 21.2 (C-5'); HRMS Calcd for $C_{11}H_{13}N_5O_3$: $(M+H)^+$ 264.1097. Found: 264.1098.

Following the same procedure, compound **23e** (70 mg, 0.13 mmol) gave, after an additional washing of the residue with CH_2Cl_2 (3 mL), nucleoside **11c** as a white powder (24 mg, 0.09 mmol, 71%).

 $(1'S^*, 2'S^*, 5'R^*, 6'S^*)$ -7-(6'-Hydroxymethyl-3'-oxabicyclo-[3.1.0]hex-2'-yl)guanine (11d). Following the same procedure as for the preparation of 11c, compound 23d (54 mg, 0.15 mmol) gave, after one week stirring, nucleoside 11d as a white powder (30 mg, 0.11 mmol, 73%): mp $312^{\circ}C$ dec. (MeOH/H₂O); IR (KBr, cm⁻¹) 3452, 1683, 1649; ¹H NMR (DMSOd₆) δ 10.87 (br s, 1H, H-1), 8.11 (s, 1H, H-8), 6.36 (s, 1H, H-2'), 6.14 (br s, 2H, NH₂), 4.60 (t, 1H, OH, J=5.4 Hz), 4.06 (dd, 1H, H-4', J=8.9, 2.0 Hz), 3.87 (d, 1H, H-4", J=8.9 Hz), 3.52 (dd, 2H, CH₂OH, J=7.3, 5.4 Hz), 2.16–2.10 (m, 2H, H-1['] and H-5[']), 1.33 (m, 1H, H-6'); ¹³C NMR (DMSOd₆) δ 159.8 (quat C), 154.6 (quat C), 152.8 (quat C), 140.7 (C-8), 107.4 (C-5), 84.2 (C-2'), 66.9 (C-4'), 54.7 (CH₂OH), 25.8 (C-1'), 21.7 (C-6'), 20.8 (C-5'); HRMS Calcd for C₁₁H₁₃N₅O₃: (M+H)⁺ 264.1097. Found: 264.1092.

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